

Dream . Challenge . Succeed

CENTER OF EXCELLENCE IN REPRODUCTIVE SCIENCES

12 March, 2007

TO:

Dr. Cleophus Price, Assistant Dean, Graduate Academic Studies

FROM:

Dr. Doris J. Baker, Director, Reproductive Laboratory Science Certificate

SUBJECT:

Review of the Reproductive Laboratory Science Graduate Certificate

Requested Summary for the Graduate Certificate in Reproductive Laboratory Science Summary

1. Introduction: The Graduate Certificate in Reproductive Laboratory Science (RLS) prepares graduates for entry level technologist positions in assisted reproductive technology (ART) laboratories and related fields in research, industry and marketing. The curriculum consists of 12-14 hours in RLS courses, including two credit hours of clinical practica in clinical andrology and embryology laboratories. The Graduate Certificate may be completed in approximately seven months of study. In addition to the RLS core faculty, lecturers throughout the United States contribute to the curriculum, instructing in their area of expertise. These professionals are ART and clinical laboratorians, laboratory managers, researchers, ethicists, attorneys and policy analysts. Directors and supervisors in andrology laboratories and ART laboratories, appointed as clinical faculty by the University of Kentucky, supervise RLS students in the clinical practica.

2. Faculty:

Core faculty:

- ➤ Doris J. Baker, Ph.D., HCLD(ABB), MT(ASCP), Professor and Director, Division of Clinical and Reproductive Sciences, Director, Graduate programs in Reproductive Laboratory Science, University of Kentucky (Full member of Graduate Faculty).
- Philip Bridges, Ph.D., Assistant Research Professor, Reproductive Laboratory Science Programs, University of Kentucky.

- ➤ Kim Campbell, M.S., MT(ASCP), Adjunct Assistant Professor and Medical Instructional Specialist, Education and Laboratory Coordinator, Reproductive Laboratory Sciences programs, University of Kentucky, Damodaran Chendil, Ph.D., Assistant Research Professor, Reproductive Laboratory Science Programs, University of Kentucky.
- Linda S. Gorman, Ph.D., MT(ASCP), Associate Professor and Director of Graduate Studies, Division of Clinical and Reproductive Sciences, (Full member of Graduate School).
- Chemyong (Jay) Ko, Ph.D., Associate Professor, Division of Clinical and Reproductive Sciences, University of Kentucky, (Associate Member of UK Graduate Faculty)
- ➤ Oliver R. Oakley, Ph.D., Assistant Professor, Reproductive Laboratory Science Programs, University of Kentucky. (Associate member of UK Graduate School)
- ➤ Julie A. Ribes, M.D., Ph.D., Associate Professor Pathology and Laboratory Medicine, University of Kentucky College of Medicine, Associate Director of Hospital Laboratories, Director of Clinical Microbiology, Assistant Director of Special Chemistry, Infectious Disease Serology; Medical Director, CLS and RLS, Division of Clinical and Reproductive Sciences, University of Kentucky, (Associate member of UK Graduate faculty)
- ➤ Jeannine Witmyer, Ph.D. Supervisor, Andrology and Cryobiology, Boston IVF, Waltham, MA; Adjunct Assistant Professor, Division of Clinical and Reproductive Sciences, University of Kentucky, (Associate Member of UK Graduate Faculty)
- Program Lecturers and Clinical Faculty lists attached
- 3. Admissions: Applicants must:
 - Meet all requirements for admission to the Graduate School at the University of Kentucky, including minimum GPA required for postbaccalaureate status
 - ➤ Hold a bachelor's degree in Clinical Laboratory Sciences (Medical Technology) OR a Bachelor's degree in science with acceptable laboratory experience
 - Complete the RLS application process, including providing 3 professional reference letters
 - Interview may be required (determined by the RLS Admissions Committee)
- 4. **Current requirements for completion** of the RLS Graduate Certificate includes successful completion of the following didactic/laboratory and clinical practica courses:
 - CSC 615 Reproductive Laboratory Science (1 credit hour/distributive learning)
 - CSC 616 Andrology (1 credit hour/distributive learning)
 - CSC 617 Reproductive Microbiology & Immunology (1 credit hour/distributive learning)

- ➤ CSC 528 Laboratory Techniques (2 credit hours; hands-on laboratory course; required for all students not having acceptable laboratory experience prior to entry into the program
- CSC 618, Laboratories in Andrology, Reproductive Microbiology & Immunology (1 credit hour; hands-on laboratory class)
- CSC 621 Embryology & ART (3 credit hours; lecture and laboratory course
- CSC 624 Gamete & Embryo Cryopreservation (1 credit hour; lecture and laboratory course
- ➤ 625 Mgt, Policy, Ethical & Legal Issues in ART (2 credit hours)
- ➤ 626 Andrology Clinical Practicum (1 credit hour; clinical practicum in accredited ART [assisted reproductive technology] laboratory); program completion requires successful completion of the andrology checklist (attached)
- ➤ CSC 627 ART Clinical Practicum (1 credit hour clinical practicum in accredited ART [assisted reproductive technology] laboratory); program completion requires successful completion of the ART/cryobiology checklist (attached)
- 5. Rationale for program continuation: The RLS Graduate Certificate should continue to be available for students wishing to enter the field of ART that already hold an acceptable degree(s). For example, an individual receiving the graduate certificate in 2006 is a UK faculty member with a Ph.D. in reproductive physiology, who wished to learn the clinical aspects of her field to expand instructional and research commitments. All RLS Graduate Certificate courses are included as part of the RLS track in the Clinical Sciences Master of Science degree. As a result additional resources, including personnel, are not required to accommodate certificate candidates.
- 6. **Advertising**: Brochure and copy of web page attached.
- 7. **Certificate**: Copy of certificate awarded to graduates is attached.

Note: An additional student has completed the RLS Graduate Certificate: Karen McDowell, Ph.D. (412-78-7832), 2006-05-07.

Lecturers

Paul Bachner, M.D. Chairman and Professor, Department of Pathology & Laboratory Medicine, University of Kentucky Chandler Medical Center, Lexington, KY; Past-President, College of American Pathologists.

Erica Behnke, Ph.D., HCLD(ABB), Erica J. Behnke, Ph.D., HCLD (ABB) Laboratory Director, Kettering Reproductive Medicine, Kettering, OH and Regional Commissioner, Reproductive Laboratory Accreditation Program, College of American Pathologists, Chicago, Illinois.

Maria Bertero, M.D., HCLD (ABB), Laboratory Director, Fertility Institute, Brooklyn, NY.

Eric Dorman, President, Embryo Tech, Wilmington, MA

Philip Campbell, M.S., MT (ASCP), Associate Professor, Eastern Kentucky University, Richmond, KY

Thomas Curry, Ph.D., Professor and , Department of Obstetrics and Gynecology, College Medicine, University of Kentucky.

Nanette Elster, J.D., M.P.H., Vice-President, Spence & Elster, P.C., Lincolnshire, IL; Adjunct Faculty, School for New Learning and College of Law; Adjunct Faculty, University of Illinois at Chicago School of Public Health

Melanie Freeman, Ph.D., CLDir (NCA), Director, Embryology Laboratory, Nashville Fertility Center, Nashville, TN

Deborah French, BHS, PA-C, MT (ASCP), Andrology Supervisor, Kentucky Fertility and Gynecology, Lexington, KY

Dayong Gao, Ph.D., Professoof Mechanical Engineering/Biomedical Engineering, University of Washington, Seattle, WA.

Robert Homm, M.D., FACOG, Fertility & Endocrine Associates, Louisville, KY

Lothar Jennes, Ph.D., Professor, Department of Anatomy & Neurology, University of Kentucky

Karen McDowell, Ph.D., Associate Professor, Department Of Veterinary Medicine, University of Kentucky

Sue Overman, M.S., MT (ASCP), Supervisor, Clinical Microbiology, University of Kentucky Medical Center

Ok-Kyong Park-Sarge, Associate Professor, Department of Physiology, College of Medicine, University of Kentucky

Patricia Payne, Ph.D., MT (ASCP), CLS (NCA), Research Participation Program, Training & Curriculum Services Division, Office of Workforce and Career Development, Centers for Disease Control and Prevention (CDC)

Douglas Powers, Ph.D., HCLD(ABB), Scientific Director, Boston IVF, Brookline, MA, Professor, Department of Biology, Boston College, Chestnut Hill, MA & Adjunct faculty, Harvard Medical School.

George Veloudis, D.O., Reproductive Endocrinologist; Kentucky Fertility and Gynecology, Paris & Lexington, KY; Voluntary faculty for Physician Assistant Program, University of Kentucky

Clinical Faculty

Marlane Angle, Ph.D., HCLD (ABB), Obstetrics and Gynecology University of Arkansas for Medical Sciences, Little Rock, AR Assistant Professor & Director, Arkansas Reproductive Technology Laboratory Department of Ob-Gyn, University of Arkansas for Medical Studies, Little Rock, Arkansas

Bill Baird, Ph.D., HCLD(ABB), Reproductive Diagnostics, Inc. & Cryobiology, Inc., Columbus, OH; In Vitro Fertilization Lab, Akron, OH, Pittsburgh Cryobank of Reproductive Diagnostics, Pittsburgh, PA and In Vitro Fertilization Lab, Kettering, OH

Pedro Beauchamp, M.D., Medical Director, Pedro Beauchamp Reproductive Endocrinology & Infertility, Puerto Rico

Erica Behnke, Ph.D., HCLD(ABB), Erica J. Behnke, Ph.D., HCLD (ABB) Laboratory Director, Kettering Reproductive Medicine, Kettering, OH and Regional Commissioner, Reproductive Laboratory Accreditation Program, College of American Pathologists, Chicago, Illinois.

Barry Behr, Ph.D., HCLD(ABB), Associate Director, Reproductive Endocrinology & Infertility, Stanford University Medical Center, Stanford, CA

Maria Bertero, M.D., HCLD (ABB), Director, Fertility Institute, Brooklyn, NY

Catherine Cowart, M.D., Director, Reproductive Endocrinology and Infertility, Clearwater, FL

Nina Desai, Ph.D., HCLD (ABB), Laboratory Director, Cleveland Clinic Fertility Center, Beachwood, OH

Barbara Estes, B.S., MT(ASCP), CLS(NCA), Supervisor, In Vitro Fertilization/Andrology Laboratory, Miami Valley Hospital, Dayton, Ohio

Deborah French, BHS, PA-C, MT (ASCP), Kentucky Fertility and Gynecology, Paris & Lexington, KY;

Marina Gavakharia, M.D., Ph.D., HCLD(ABB), Laboratory Director, Fertility Physicians of Northern California, San Jose, CA

Dolores Lamb, Ph.D., HCLD (ABB), Associate Professor or Urology & Cell Biology and Director, Laboratory for Male Research & Testing, Baylor College of Medicine, Houston, TX Department of Ob-Gyn, University of Arkansas for Medical Studies, Little Rock, Arkansas

Karen McDowell, Ph.D., Associate Professor, Department of Veterinary Medicine, University of Kentucky

Terry Olar, Ph.D., HCLD (ABB), Director, Center for Reproductive Studies, Christ Hospital, Cincinnati, OH

Margaret Papadakis, Ph.D., HCLD (ABB), Director, Reproductive Biology Laboratories, Department of Ob-Gyn, Carolina Medial Center, Charlotte NC

Charles Sims, M.D., Medical Director, Fertility Center of California, Los Angeles, CA

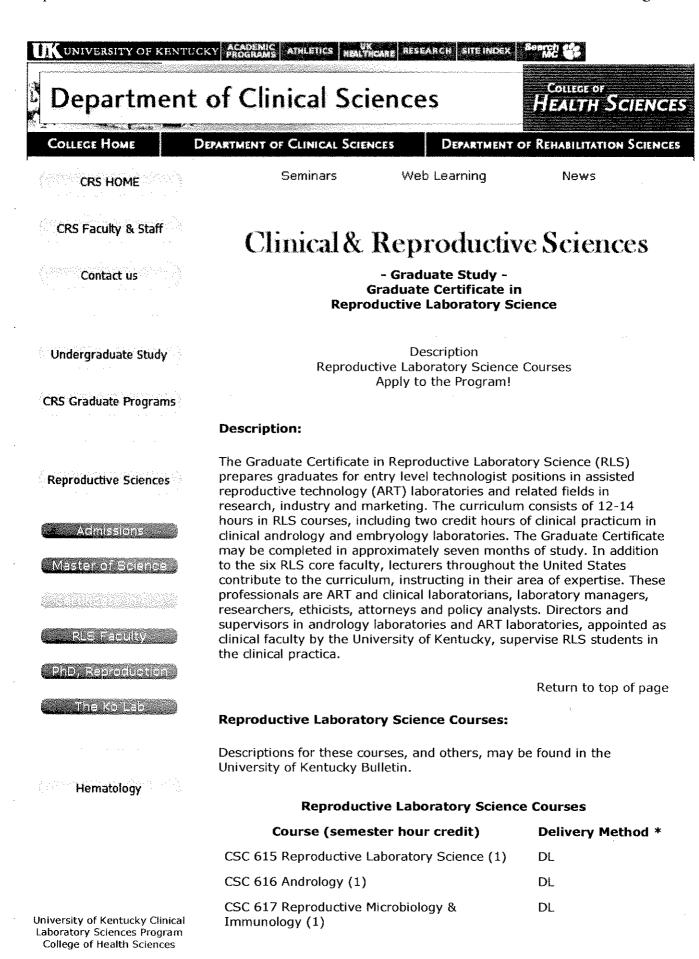
Timothy Smith, Ph.D., HCLD (ABB), Laboratory Director, North Hudson IVF, Englewood Cliffs, NJ

Samer Soubra, B.S., CLS, Embryologist & Laboratory Supervisor, Reproductive Partners Medical Group, Inc. Long Beach, CA

Andras Szell, Ph.D., HCLD (ABB), Laboratory Director, Alta Bates, IVF, Berkeley, CA

Carolyn Walters, ARNP, Advance Practice Nurse Practitioner, (Ob-Gyn), Utica Park Clinic, Tulsa, OK

Jeannine Witmyer, Ph.D., HCLD(ABB), Adjunct Faculty, Reproductive Laboratory Sciences Graduate Programs, University of Kentucky; Supervsior of Andrology and Cryobiology, Boston IVF, Waltham, MA.



NOTICE: Some Web sites to
which these materials provide
links for the convenience of
users are not managed by the
University of Kentucky. The
University does not review,
control or take responsibility

for the contents of those sites.

Lexington, KY 40536-0200

CSC 528 Laboratory Techniques (2)
Students not having an acceptable laboratory
background will also be required to complete
CSC 528

M, 1 week of 8 wk Summer Session

CSC 618 Labs in Andrology, Reproductive Microbiology & Immunology (1)

M, 1 week of Fall Semester

CSC 621 Embryology & ART (3)

Fall Semester

CSC 624 Gamete & Embryo Cryopreservation (1) Fall Semester

CSC 625 Mgt, Policy, Ethical & Legal Issues in

Fall Semester

ART (2)

CSC 626 Andrology Clinical Practicum (1)

1 weeks TBD

CSC 627 ART Clinical Practicum (1)

1 weeks TBD

- * DL = Distributive learning. DL courses include web-based instruction and testing, and self-paced, interactive CDs. DL courses are offered during the spring semesters to both Lexington campus students and distance learners.
- * M = Modular. Modular courses are taught at the UK campus. Classes meet for approximately 8 hours/day.
- * TBD = To Be Determined

NOTE = All Clinical practica will take place in Assisted reproduction laboratories in the U.S. under the supervision of Directors appointed as clinical faculty by the University of Kentucky.

Return to top of page

The University of Kentucky is accredited by the Commission on Colleges of the Southern Association of Colleges and Schools (1866 Southern Lane, Decatur, Georgia 30033-4097: Telephone number 404-679-4501) to award undergraduate, graduate and professional degrees.

The University of Kentucky is committed to a policy of providing educational opportunities to all qualified students regardless of economic or social status, and will not discriminate on the basis of race, color, religion, sex, marital status, beliefs, age, national origin, sexual orientation, or physical or mental disability.

NOTICE: Although every effort is made to ensure that this material is accurate and up to date, it is provided for the convenience of the user and should not be considered official. The official version of this material is available in the University of Kentucky Bulletin. The user is advised to refer to and rely upon the official version of this material when making significant decisions or judgments.

Comments to Oliver R Oakley oroakl1@uky.edu, Last Modified: March 12, 2007 Copyright @ 2007, University of Kentucky The University of Kentucky is an Equal Opportunity University Terms, Conditions & Privacy Statement

FAX: 859-323-8957

INSTRUCTIONS for ART/EMBRYOLOGY checklist:

- The checklist is comprehensive (e.g. including hormone evaluations)
- Respond only to applicable sections
- Place tally mark for each experience or assay so a total can be compiled later
- Please complete the comments section
- A general summary may included in the Comments Section on the last page
- Supervisors sign the checklist

ALL comments and suggestions are appreciated.

Doris J. Baker, Ph.D., HCLD(AAB), MT(ASCP)

dbake0@uky.edu Telephone: 859-323-1100 ext. 80854

ART/EMBRYOLOGY CHECKLIST

	Policy, Rules, Procedures Explained by Instructor	Methodology	Observed by Student	Performed under Supervision	Performed Independently	Comments
Media Preparation						
a. Oocyte Collection Media medium						
b. Culture medium	·					
c. Sperm capacitation/prep medium						
d. Oil						
e. Transfer medium						
f. OTHER						
IVF						
Patient identification						
a. Releases						
b. Site specific paperwork (including Chain of Custody)						
Oocyte retrieval (patient or donor)						
a. Identification						
b. Assessment						

_2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

	Policy, Rules.	Methodology	Observed			Comments
	Procedures Explained by Instructor		by Student u	under	Independently	
c. Site specific paperwork						
Semen Preparation						
a. Concentration/motility						
b. Recovery of motile fraction						
c. Calculations						
Insemination (partner or donor)						
a. Calculations						
b. Insemination						
c. Site specific paperwork						
ICSI						
ICSI (practice with non-fert oocytes)						
Fertilization						
a. Documentation of fertilization						
b, Recheck						
c. Reinsemination (if applicable)						

_2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

Policy. Rules, Methodology Observed Performed Comments Procedures Explained by Student under Supervision Independently Instructor	¥.			ic	k			oryo				heck	X.			
Policy. Rules, Metho Procedures Explained by Instructor														***************************************		
	d. Site-specific paperwork	Embryo culture	a. Scoring	b. Preimplantation Genetic Diagnosis	c. Site specific paperwork	Embryo Transfer	a. Scoring	b. Sorting for transfer vs cryo	c. Assisted hatching	d. Patient identification verification	e. Loading catheter	f. Post-transfer catheter check	g. Site-specific paperwork	GIFT/ZIFT	a. Patient identification	h Cite ensoriffe noncontract

_2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

	Policy, Rules, Procedures Explained by Instructor	Methodology	Observed l	Performed under Supervision	Performed Independently	Comments
including Chain of Custody						
c. Semen preparation (GIFTonly)						
 d. Oocyte/embryo assessment & sorting 			•			
e. Tubal transfer						
Cryopreservation of embryos/oocytes (patient or donor)						
a. Informed consent						
b. Specimen identification						
c. Assessment of quality						
d. Specimen freezing						
e. Site specific logging/documentation (including <i>Chain of Custody</i>)						
Thawing embryos/oocytes (patient or donor)						
a. Informed consent						
b. Specimen identification	***************************************					

2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

	Policy. Rules, Procedures Explained by Instructor	Methodology	Observed by Student	Performed under Supervision	Performed Independently	Comments
c. Thawing						
d. Post-thaw quality assessment						
e. Site specific logging/documentation (including <i>Chain of Custody</i>)						
PRACTICE/Mouse Model						
a. Embryo thaw						
b. Culture						
c. Scoring for development to blastocyst						
d. Calculate percent survival						
PRACTICE/Non-fertilized oocytes						
a. Handling						
1. Transfer between dishes						
2. Other						
Instrumentation	**************************************					
a. Calibration					-	
b. Preventative maintenance						

2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

	Policy, Rules, Methodology Procedures Explained by Instructor	Methodology	Observed by Student	Observed Performed by Student under Supervision	Performed Independently	Comments
Perform Quality Control						
a. Daily						
b. Weekly						
c. Periodic						
Participate in Proficiency Testing (if applicable)						

Participate in Site-specific Quality Assurance program when applicable: List examples

Please list any additional procedures/experiences

1					
					1
	ıts:			(s)e	
Student	Comments:			Signature(s)_	
Stu	Coi			Sig	

2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

ANDROLOGY CLINICAL PRACTICUM

STUDENT	
CLINICAL SITE	
CLINICAL PRACTICA DATES	
PRIMARY SUPERVISOR	

INSTRUCTIONS for ANDROLOGY checklists:

- The checklist is comprehensive (e.g. including hormone evaluations)
- Respond only to applicable sections
- Place tally mark for each experience or assay so a total can be compiled later
- Please complete the comments section
- A general summary may included in the Comments Section on the last page
- Supervisors sign the checklist ALL comments and suggestions are appreciated.

Doris J. Baker, Ph.D., HCLD(AAB), MT(ASCP)

dbake0@uky.edu Telephone: 859-323-1100 ext. 80854

FAX: 859-323-8957

ANDROLOGY CHECKLIST

	1000	A CONTRACT OF THE PERSON OF TH				
	Policy, Rules, Procedures Explained by Instructor	Methodology (e.g. Makier vs. CASA)	Observed by Student	Performed under Supervision	Performed Independently	Comments
Patient Instruction for Semen Analysis						
a. Collection						
b. Transport						
Specimen Processing						
a. Patient information data						
b. Specimen handling and logging					•	
Perform Semen Analysis						
a. Macroscopic Assessment	300				Programme and the second secon	
1. Coagulum/liquefaction						
The same and the s						

2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

									,
Comments									
Performed Independently									
Performed under Supervision									
Observed by Student									
Methodology (e.g. Makler vs. CASA)									
Policy, Rules, Procedures Explained by Instructor									
	2. Volume	3. Viscosity	4. Appearance	5. pH	b. Microscopic Assessment	I. Initial Assessment of Quality (e.g. debris, clumping, epithelial, rbcs other)	2. Quantitative Motility	3. Agglutination % (if indicated on initial assessment)	4. Vitality (if indicated on motility)

_2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

	Т	I						I	
Comments									
Performed Independently									
Performed under Supervision									
Observed by Student									
Methodology (e.g. Makler vs. CASA)									
Policy, Rules, Procedures Explained by Instructor							E La Calabatan de la Calabatan		
	5. Concentration	6. Total sperm count	7. Make and stain morphology slides	8. Read morphology	9. Round cell assessment	10. OTHER	Leukocyte Detection	a. Staining (Peroxidase); Leukostain	b. Immunocytochemistry

2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

Comments									
Performed Independently									
Performed under Supervision									
Observed by Student									
Methodology (e.g. Makler vs. CASA)									
Policy, Rules, Procedures Explained by Instructor									
	с. ОТНЕК	Semen Microbiology	a. Gram stain	b. Aerobic cultures	c. Anaerobic cultures	d. Mycoplasma/Ureaplasma	e. OTHER (e.g. Chlamydia)	Semen Biochemistry	a. Acid phosphatase

2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

[<u> </u>			[]	1
Comments									
Performed Independently		:							
Performed under Supervision									
Observed by Student									
Methodology (e.g. Makler vs. CASA)									
Policy, Rules, Procedures Explained by Instructor					***************************************				
	b . Alpha-glucosidase	c. Citric acid	d. Fructose	e. Zinc	f. OTHER	Sperm Function	a. Acrosin	b. Hypo-osmotic swelling test	c. Mannose binding assay

_2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

_2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

Comments									
Performed Independently									
Performed under Supervision									
Observed by Student									
Methodology (e.g. Makler vs. CASA)									
Policy, Rules, Procedures Explained by Instructor									
	a. Sperm-Cervical Mucus Interaction Test	b. Sperm-Cervical Mucus Cross Test	c. OTHER	Hormone Evaluations	a. Estradiol	b. FSH	с. І.Н	d. Progesterone	e. Prolactin

2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

Comments								MATERIAL PROPERTY.	
Performed Independently									
Performed under Supervision									
Observed by Student								-	
Methodology (e.g. Makler vs. CASA)									
Policy, Rufes, Procedures Explained by Instructor									
			_		ation		ttion	Semen	
	'ne		ervation lonor)	consent	ı identific	rfreezing	ific	reserved lonor)	consent
	f. Testosterone	g. OTHER	Semen Cryopreservation (patient or donor)	a. Informed consent	b. Specimen identification	c. Specimen freezing	d. Site-specific logging/documentation	Thawing Cryopreserved Semen (patient or donor)	a. Informed consent
	(4-i	oio	Semen (a. I	, .d	, ,	ਚ	Thawin; (pa	rej esi

2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

b. Specimen identification c. Thawing d. Sperm survival rate documentation e. Site-specific logging/documentation including Chain of Clustody b. Specimen preparation (wash or concentration) c. Site-specific logging/documentation including c. Site-specific logging/documentation including Chain of Clustody) Andrology Instrumentation		Policy, Rules, Procedures Explained by Instructor	Methodology (e.g. Makler vs. CASA)	Observed by Student	Performed under Supervision	Performed Independently	Comments	1
c. Thawing d. Sperm survival rate documentation e. Site-specific logging/documentation including Chain of Custody Semen Preparation for Insemination a. Specimen identification b. Specimen preparation (wash or concentration) c. Site-specific logging/documentation including Chain of Custody) Andrology Instrumentation	b. Specimen identification							
e. Site-specific logging documentation a. Specimen preparation (wash or concentration) c. Site-specific logging documentation including c. Site-specific logging documentation including c. Site-specific logging documentation including Chain of Custody) Andrology Instrumentation	c. Thawing							
e. Site-specific logging/documentation including Chain of Custody: Semen Preparation for Insemination a. Specimen identification b. Specimen preparation (wash or concentration) c. Site-specific logging/documentation including Chain of Custody) Andrology Instrumentation	d. Sperm survival rate documentation							
a. Specimen identification b. Specimen preparation (wash or concentration) c. Site-specific logging/documentation including Chain of Custody) Andrology Instrumentation	e. Site-specific logging/documentation including Chain of Custody							
a. Specimen identification b. Specimen preparation (wash or concentration) c. Site-specific logging/documentation including Chain of Custody) Andrology Instrumentation	Semen Preparation for Insemination							
b. Specimen preparation (wash or concentration) c. Site-specific logging/documentation including Chain of Custody) Andrology Instrumentation	a. Specimen identification							т
c. Site-specific logging/documentation including Chain of Custody) Andrology Instrumentation	b. Specimen preparation (wash or concentration)							1
Andrology Instrumentation	c. Site-specific logging/documentation including Chain of Custody)							
	Andrology Instrumentation							

2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

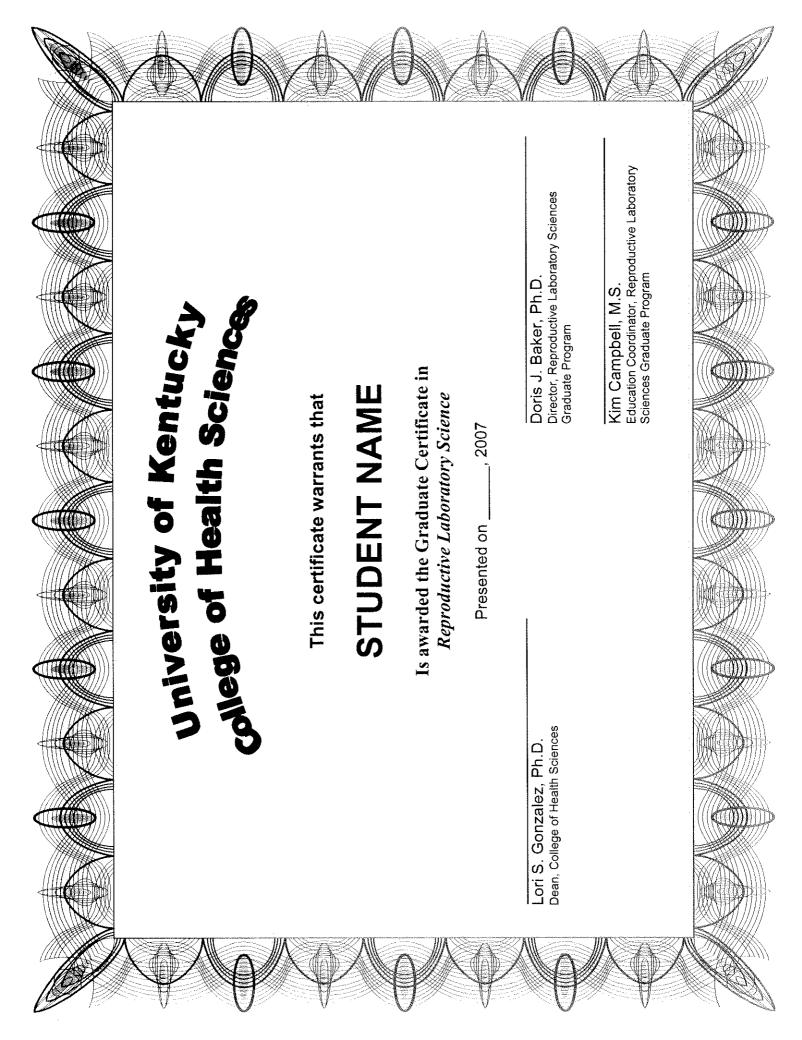
ed Comments mtly							
Performed Independently n							
Performed under Supervision							
Observed by Student							
Methodology (e.g. Makler vs. CASA)							
Policy, Rules, Procedures Explained by Instructor							
	a. Calibration	b. Preventative maintenance	Perform Quality Control	a. Daily	b. Weekly	c. Periodic	Participate in Proficiency Testing (if applicable)

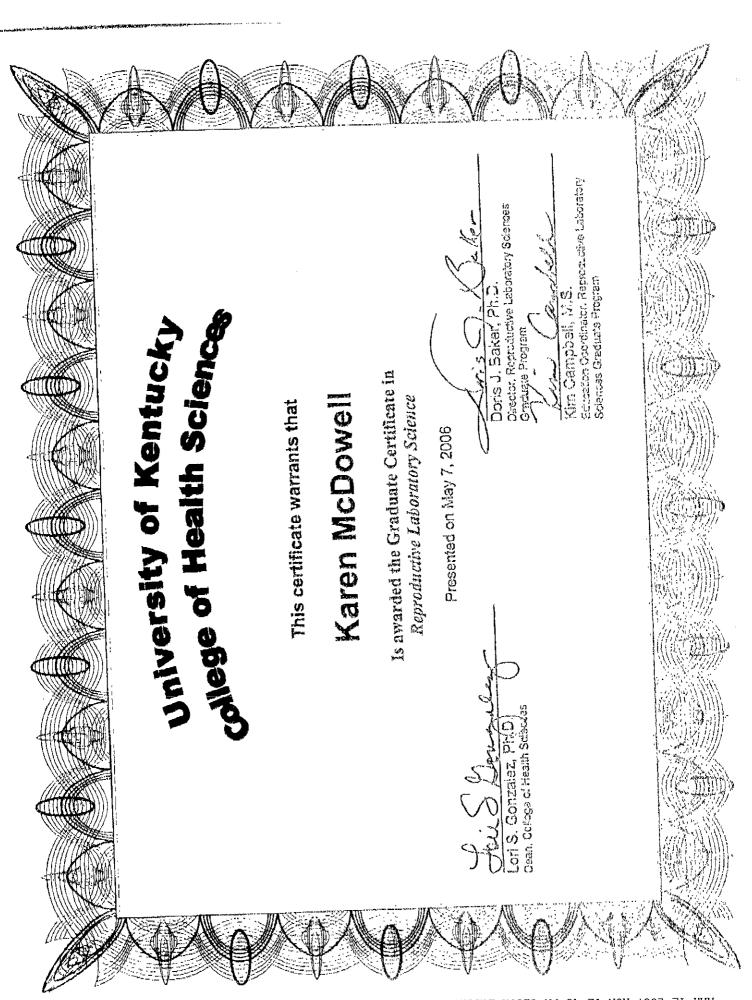
_2006 Doris J. Baker, Ph.D., Kim Campbell, M.S. & Jeannine Witmyer, Ph.D.

Student				
Comments:				
Signatura(c)				
Signature(s)_				
		· · · · · · · · · · · · · · · · · · ·		
		~	 	

ART/EMBRYOLOGY CLINICAL PRACTICUM

STUDENT	
CLINICAL SITE	
CLINICAL PRACTICA DATES	
PRIMARY SUPERVISOR	





Trans Number	Date Received	Date Assigned	Date to Council	Sub Comm	College
0607-P015					
0607-P016	03/05/07	03/05/07		HUM/Jensen	Music
0007010	8/23	1723		Envir	DIVINE
0607-P017	8/23	3/23	•	DeVI	e108
0607-P018	3/23			Colle	ge T
0607-P019	3/53			En Viv De VI Colle	llesc
0607-P020	3/23			500	19/7
0607-P021	3/23			MS	1917 10
0607-P022	3/23			Re	Prod
0607-P023	<u> </u>				
0607-P024					
0607-P025					
0607-P026					
0607-P027					
0607-P028			~~~~		
0607-P029					
0607-P030					